

# Update on the Molecular Pathogenesis of Pancreatic Tumors Other than Common Ductal Adenocarcinoma

D. Antonello<sup>a</sup> S. Gobbo<sup>a</sup> V. Corbo<sup>a</sup> B. Sipos<sup>b</sup> N.R. Lemoine<sup>c</sup> A. Scarpa<sup>a</sup>

<sup>a</sup>Dipartimento di Patologia, Università di Verona, Verona, Italy; <sup>b</sup>Department of Pathology, University of Kiel, Kiel, Germany, and <sup>c</sup>Institute of Cancer and CR-UK Clinical Centre, Barts and The London School of Medicine, London, UK

## Key Words

Pancreas · Pathogenesis · Tumor

## Abstract

**Purpose:** Although ductal adenocarcinoma is the most common and well known pancreatic tumor type, other distinct epithelial neoplasms affecting the pancreas that show different symptoms, biological behaviors and outcomes are becoming more frequently recognized and documented. Pancreatic epithelial tumors may be separated into ductal and nonductal neoplasms. The former group includes pancreatic ductal adenocarcinoma, intraductal papillary-mucinous tumor, mucinous cystic tumor and serous cystic tumor. The latter group includes pancreatic endocrine tumor, pancreatic acinar cell carcinoma, pancreatoblastoma and solid-pseudopapillary tumor. The aim of this review is to summarize recently acquired knowledge regarding the molecular characterization of these uncommon pancreatic epithelial neoplasms. **Recent Findings:** Molecular studies of uncommon pancreatic epithelial tumors suggest that the different morphological entities are associated with distinct molecular profiles, highlighting the involvement of different molecular pathways leading to the development of each subtype of pancreatic neoplasm. **Conclusion:** The correct classification of rare pancreatic epithelial tumors and the

identification of their characteristic molecular aspects is the fundamental starting point in identifying novel diagnostic molecular tools and new targets for innovative therapeutic strategies.

Copyright © 2008 S. Karger AG, Basel and IAP

## Introduction

Although ductal adenocarcinoma is the most common tumor affecting the pancreas [1], other distinct epithelial neoplasms that show a wide range of symptoms, biological behaviors and outcomes are becoming increasingly recognized and characterized [2].

Pancreatic epithelial neoplasms develop from three main cell lineages. Most pancreatic neoplasms have ductal differentiation, including the pancreatic ductal adenocarcinoma (PDAC), as well as intraductal papillary-mucinous tumor (IPMT), mucinous cystic tumor (MCT) and serous cystic tumor (SCT). The nonductal pancreatic epithelial neoplasms develop from either the islet of Langerhans' cells giving rise to pancreatic endocrine tumor (PET) or from acinar cells as pancreatic acinar cell carcinoma (PACC), pancreatoblastoma (PBL) and solid-pseudopapillary tumor (SPT). The molecular mechanisms underlying the pathogenesis of pancreatic epithe-

lial neoplasms are poorly understood [2]. The aim of this paper is to collect and report the recent findings in the molecular characterization of uncommon pancreatic epithelial neoplasms.

### Intraductal Papillary-Mucinous Tumor

IPMT is a noninvasive, mucin-producing, epithelial neoplasm, predominantly arranged in papillae that grow inside the ductal system. They are morphologically classified as adenoma, borderline, or noninvasive carcinoma according to the degree of the cellular atypia [3, 4]. IPMT may show an invasive neoplastic component which leads to a diagnosis of 'IPMT-associated invasive adenocarcinoma' characterized by a poorer prognosis [3, 5–8]. With regard to the molecular pathogenesis, IPMT shows *KRAS*-activating mutations as an early event that increases with histological severity of IPMT. A recent report shows that DNA damage checkpoint activation due to *CHK2* inactivation or *TP53* mutation occurs in the early stage of IPMT and prevents their progression thus contributing to the carcinogenesis of IPMT [9 – it illuminates the role of *TP53* pathway in the IPMT development]. *AKT/PKB* and *HER2/EGFR* activation is reported in a large fraction of IPMT; expression of *CDKN2A/P16* is frequently lost, suggesting a correlation with the hypermethylation of the promoter region of *P16*, more frequently detected in high-grade tumors; some IPMT show abrogation of *TP53*, especially those with severe/high grade atypia [10 – an excellent summary of IPMT molecular pathogenesis, 11–16]. Despite the frequent hemizygous or homozygous deletions of chromosome 18q, *SMAD4* is completely retained in IPMT [17, 18]. Mutation of *STK11/LKB1*, a Peutz-Jeghers syndrome gene, and abrogation of the expression of *DUSP6/MKP-3*, a gene identified in the deleted region 12q21-q22, indicate the possible role of these molecules in the development of a subset of IPMT [16, 19, 20]. Aberrant hypermethylation of at least one CpG island was detected in about 80% of IPMT, with the overall number of methylated loci significantly higher in high-grade tumors. *Cyclin D2*, *TFPI-2* and *SOCS-1* have been reported as aberrantly methylated in IPMT [21 – detailed descriptions of relevant epigenetic alterations in IPMT]. Global gene expression analysis performed for IPMT revealed that many of the overexpressed genes in IPMT are also highly expressed in PDAC. Gene expression profiles showed upregulation of members of the trefoil factor family (*TFF1* and *TFF3*), *CLD4*, *CXCR4*, *S100A4*, and *Mesothelin*, some of which have been suggested as playing a

role in the progression to the invasive form of IPMT [22–24]; among underexpressed genes in IPMT, *CDKN1C/P57KIP2* has been shown to be epigenetically downregulated [24]. Recent investigations suggest that the *SHH* pathway might play a role during tumorigenesis of IPMT and that *SHH* measurement of pancreatic juice may provide some advantages in the treatment or follow-up of a subset of patients with IPMT [25 – an outstanding survey of *SHH* pathway involvement in IPMT with its possible clinical implications, 26 – reveals *SHH* pathway role in various pancreatic tumors, 27 – an immunohistochemical study of *SHH* expression in IPMT confirming the role of the pathway in this neoplasm]. *Fascin* expression was found to be significantly higher in borderline neoplasms and carcinomas than in adenomas, suggesting that *Fascin* overexpression is involved in the progression of IPMT. *Fascin* could become a new therapeutic target for the inhibition of their progression [28 – identification of *Fascin* as possible prognostic marker in IPMT]. *PIK3CA* mutations in 11% of IPMT have been reported, providing evidence that the oncogenic properties of *PIK3CA* contribute to the tumorigenesis of IPMT [29 – a survey of types and numbers of *PIK3CA* mutations in IPMT]. Recent results suggest that *HTERT* expression in epithelial cells indicates malignant transformation in IPMT and immunohistochemical detection of *HTERT* in cells derived from pancreatic juice may represent a powerful diagnostic tool [30 – identification of *HTERT* as possible marker of malignant progression in IPMT]. *MUC4* and *MUC5AC* have been recently revealed as potential markers in distinguishing more aggressive IPMNs from less malignant ones [31 – it suggests the use of *MUC4* and *MUC5AC* to distinguish malignant from benign IPMT lesions].

### Mucinous Cystic Tumor

MCT are cyst-forming and mucin-producing epithelial neoplasms characteristically lying on an ovarian-like stroma and showing various degrees of cytoarchitectural atypia. MCT surgically resected and diagnosed as noninvasive neoplasms have an excellent prognosis. The rare cases of recurrence and tumor-related death are always due to deeply invasive MCT [32, 33].

Since MCT usually contain a small number of neoplastic cells, molecular studies are difficult. Epithelial cells of MCT express *Cytokeratins*, *Epithelial Membrane Antigen*, *Carcinoembryonic Antigen* and some *Mucins* [3, 34 – an outstanding comprehensive review of MCT pathogenesis, 35], while the immunoexpression of an  $\alpha$ -

*integrin* subunit has been reported to be independently correlated to the malignant potential [34, 36].

The frequencies of abnormal *TP53* accumulation and *KRAS* gene point mutations vary among published reports but have been consistently reported to increase with the grade of cell atypia. Hypermethylation of *P14* and *P16* was found in benign and borderline cases of MCT [37–39]. A recent study shows that MCT arises from the concomitant expression of *KRAS* (G12D) and haploinsufficiency of the *SMAD4/DPC4* tumor suppressor gene, culminating in invasive ductal adenocarcinoma with a progression scheme analogous to the classical sequence from intraductal neoplasia to ductal adenocarcinoma [40 – it illuminates the significance of the sequence and the context of critical mutations acquisition in determining the ensuing pathology]. MCT exhibits an autocrine regulation of the *SHH* pathway, suggesting a role in its tumorigenesis [26]. Expression profiling studies in MCT have revealed the overexpression of *S100P*, *Prostate stem cell antigen*, *Jagged1*, *HES1*, *STK6/STK15*, *Cathepsin E*, and *Pepsinogen C* by the neoplastic epithelium. *Jagged1* and *HES1* are members of the Notch signal pathway, which has been observed in several human cancers. The same analysis revealed the overexpression in the ovarian-type stroma of several genes involved in estrogen metabolism, including *Steroidogenic Acute Regulatory* and *Estrogen Receptor 1* genes [34, 41], suggesting that luteinized cells in the ovarian-type stroma of pancreatic MCT are capable of steroidogenesis [42]. MCC1 is the first cell line derived from a noninvasive pancreatic MCT and is expected to contribute to the detection of important molecular features of this neoplasm [43].

### Serous Cystic Tumor

The group of SCT includes three subtypes: serous microcystic adenoma, serous oligocystic ill-demarcated adenoma and von Hippel-Lindau (VHL)-associated SCTs. Since these subtypes show different biological characteristics, they are thought to represent different entities. SCT exhibiting locally invasive growth pattern and apparent distant metastases are limited to isolated case reports [2, 44–47].

Despite their biological differences, the different subtypes of SCT seem to share common morphological aspects and immunoprofiles. SCT is characterized by immunoeexpression of *Cytokeratins* and *Neuron-specific Enolase* whereas *Vimentin* and *Synaptophysin* are not found. A common centroacinar origin is also supported

by the finding that a number of SCT show *MUC1* and *MUC6* immunoeexpression similar to normal pancreatic centroacinar cells.  $\alpha$ -*Inhibin* and *MUC6* may be regarded as new markers for this type of pancreatic tumor [2, 44, 48–50].

The molecular pathogenesis of SCT does not involve *P16*, *TP53* and *SMAD4*, in contrast to PDAC [39, 51]. VHL-SCT is characterized by both LOH at chromosome 3p (which contains the *VHL* gene) and *VHL* gene germline mutation. More than 50% of serous microcystic adenomas show LOH at 10q, while only 40% have LOH at chromosome 3p and only 22% exhibit a somatic *VHL* gene mutation [52].

### Pancreatic Endocrine Tumor

Most PET are well-differentiated tumors divided into functioning PET (F-PET), a heterogeneous group of malignancies with various clinical symptoms due to hormonal hypersecretion, and nonfunctioning PET (NF-PET) without these clinical symptoms. On the basis of various morphological and biological criteria, PET are classified into benign PET, PET with uncertain malignant potential and PET showing low-grade or high-grade malignancy [53–55].

Many studies state that PET arise from distinctly different molecular pathways and are unrelated to ductal cancers [2, 51, 56–59]. To date, mutation of *MEN-1* and loss of chromosome 11q, which encompasses the region containing the *MEN1* locus, are the most common genetic alterations found in PET, with higher frequencies in gastrinoma and NF-PET than in insulinoma [60–64]. In general, the risk of metastatic spread does not appear to relate to the presence of *MEN1* mutations in sporadic PET [62].

Chromosomal analysis has identified numerous regions of chromosome losses and gains in sporadic PET and has suggested the existence of two subgroups: those showing frequent allelic imbalances and those showing low allelic imbalances [65, 66, 67 – the last of these shows the usefulness of genome-wide SNP analysis for the AI detection in PET]. The allelotype of NF-PET is, moreover, markedly different from that of ductal, acinar, or serous tumors of the pancreas, as well as from that of F-PET [52, 65, 68–70]. Moreover, recent studies reveal that the total number of genomic changes per tumor appears to be associated with both the tumor volume and the stage of the disease, indicating the accumulation of genetic alterations during tumor progression [66, 67]. These

findings point to chromosomal instability as an important mechanism associated with tumor progression. A recent study concluded that the DNA copy number status is the most sensitive and efficient marker of adverse clinical outcome in insulinoma and of potential interest in noninsulinoma PET [71 – it proves the reliability of DNA copy number status as a prognosticator to improve clinical diagnosis in insulinomas and its potential interest in noninsulinoma PET].

Studies aimed to characterize gene expression profiles of PET have been recently published, showing the involvement of various molecules that influence several biological processes such as cell-cycle progression, vascular growth, signal transduction through tyrosine kinases and cellular migration. Some of the genes identified, like *BIN1*, *Serpine10*, *BST2*, *LCK*, *IGFBP3*, *MET* and *Fibronectin*, are potential new molecular markers for the detection and treatment of these tumors [72 – it discloses a list of differentially expressed genes in a uniform set of aggressive NF-PETs and reveals a previously unknown high level of similarity between metastatic and primary lesions, 73, 74, 75 – the last of these provides an insight into PET tumorigenesis and the identification of new possible markers].

A recent investigation of the global microRNA expression patterns in normal pancreas, PET and PACC shows that a particular pattern of microRNA expression distinguishes PET from normal pancreas and PACC, suggesting that this set of microRNAs might be involved in PET tumorigenesis. This study also showed that *miR-204* is primarily expressed in insulinomas and correlates with immunohistochemical expression of insulin and that the overexpression of *miR-21* is strongly associated with both a high Ki67 proliferation index and presence of liver metastases. These results suggest that alteration in microRNA expression is related to endocrine neoplastic transformation and progression of malignancy, and might prove useful in distinguishing tumors with different clinical behavior [76].

Some factors like *VEGF-C*, *MAGE-1*, *P27/KIP1*, *Thrombomodulin* and *SRC kinases* have been described as being involved in metastatic spread of PET, thus suggesting new therapeutic approaches to decrease the malignancy of these tumors [77 – it suggests a fundamental role for *SRC Kinases* in metastatic PET, 78–81]. Other molecules recently proposed as possible targets for novel forms of therapy for PET are *CDK4*, *PDGFR- $\beta$* , *CLDN 3* and *CXCL-12* [82 – it shows ubiquitous expression of *CDK4* in PET suggesting its employment in target therapy, 83 – it suggests that new therapeutic options to inhibit the growth and spread of PET could include targeting of *PDGFR- $\beta$* ,

84 – it shows *CLD3* overexpression in PET, 85 – it illuminates angiogenesis in PET revealing *CXCL-12* as the first neoangiogenesis-associated molecule candidate].

Ploidy, sex chromosome loss and *ARHI* expression seem to be prognostic factors for disease outcome in PET [65, 86 – it suggests the use of *ARHI* expression as a new prognostic marker in PETs, 87], while expression of *Clusterin*, *Ghrelin receptor*, *Utrophin* and *Cyclin D1* did not relate to tumor aggressiveness [88–90].

Methylation studies revealed silencing of *RASSF1A*, *P16/INK4A*, *O6-MGMT*, *RAR-B* and *HMLH1* [91]. A recent paper has suggested that chromatin remodeling by histone acetylation might play a role in pancreatic endocrine cancer, as the histone-deacetylase inhibitor trichostatin A strongly inhibits cell growth of different pancreatic endocrine carcinoma cell lines [92 – it discloses mechanisms involved in growth inhibition of PET cell lines by TSA]. Finally, a new model to study genetic alterations and new therapies associated with the progression from normal cells to hyperplasia to islet cell tumors has been established by overexpression of *hTS* in murine islets [93].

#### **Pancreatic Acinar Cell Carcinoma/ Pancreatoblastoma**

PACC and PBL are rare pancreatic tumors with the features of acinar cells. PBL is a childhood tumor, whereas PACC affects adults and shows an aggressive clinical course with early metastases to the liver [94–96]. In both of these neoplasms, mutations in *KRAS* are exceedingly rare while *TP53* mutations and alterations in *P16* or *DPC4* are absent [51, 97]. Genome-wide allelotyping of these tumors has shown a high degree of allelic loss distinct from other pancreatic tumors and the involvement of chromosome 4q and 16q seems to be characteristic of this subtype [68]. Also the aberrant nuclear expression of  $\beta$ -catenin in PBL and the focal expression of *Topoisomerase II $\alpha$*  in PBL and PACC suggest that genetic pathways of these neoplasms differ from those characteristic of PDAC [98]. Elevated  $\alpha$ -fetoprotein blood concentration, a common tumor marker used to screen for hepatocellular carcinoma in high-risk patients, can also occur in PACC and PBL [99, 100, 101 – a comprehensive review of the clinical presentation, etiology, diagnosis, treatment and prognosis of PBL].

In PACC, the most common molecular alterations are the allelic loss on chromosome 11p and alteration of the *APC/ $\beta$ -catenin* pathway [102]. A study of global micro-

RNA expression in PACC showed that a common pattern of microRNA expression distinguishes this tumor from PET and normal tissue, suggesting that a set of microRNAs might be related to acinar neoplastic transformation and progression of malignancy [76].

PBL displays morphological, immunohistochemical, and clinical features that may overlap with those of PACC. Furthermore, the molecular changes are similar, showing common alteration in APC/ $\beta$ -catenin pathway, whereas no microsatellite instability or mutations of the *KRAS*, *TP53* and *DPC4* genes have been observed [103]. Moreover, PBL have been revealed to overexpress *IGF2* and to show allelic losses at chromosome 11p. Interestingly, the altered chromosomal region includes the *WT2* locus, a gene involved in other childhood tumors and in the Beckwith-Wiedemann syndrome, which may be associated with PBL [103, 104]. Morphologically, PBL is characterized by the accumulation of squamoid corpuscles which may be related to the deposition of  $\beta$ -catenin [105]. The squamoid corpuscles usually lack the features of complete squamous metaplasia and show a characteristic keratin expression [106]. *RASSF1A* promoter methylation is found in the majority of pediatric tumors, including PBL [107].

### Solid Pseudopapillary Tumor

SPT typically affects young women and generally is a low-grade malignant tumor, even if uncommon cases showing aggressive behavior have been reported. Morphologically, it is characterized by a solid and pseudopapillary architecture [32, 108–110].

SPT has a molecular pathogenesis distinct from PDAC [51, 111]. LOH on chromosome 5q22.1, mutations in exon-3 of the  $\beta$ -catenin gene and nuclear accumulation of  $\beta$ -catenin are involved in SPT tumorigenesis [111, 112 – a comprehensive survey of mutations in exon-3 of the  $\beta$ -catenin gene, nuclear accumulation of  $\beta$ -catenin, and LOH on chromosome 5q22.1 in SPT tissue, 113]. Abnormal expression of the *E-cadherin*/ $\beta$ -catenin complex may explain the cystic changes, due to the discohesive nature of the neoplastic cells and provides an additional diagnostic feature [114 – it reports changes in the expression of the principal members of the E-cadherin/catenin complex unique to SPTs, 115 – it shows that nuclear expression of  $\beta$ -catenin and loss of E-cadherin are the most reliable tests to be used in the definite diagnosis of SPT]. More accurately, it has been suggested that loss of cytoplasmic  $\beta$ -catenin protein in the cell adhesion complex results in instability of the complex, loss of *E-cadherin* in

cell membrane, and eventually dissociation of the tumor cells to form a pseudopapillary pattern [116]. The recent demonstration of aberrant nuclear localization of *E-cadherin* protein in SPT may be of diagnostic value in concert with  $\beta$ -catenin staining [117 – this study is the first demonstration of aberrant nuclear localization of *E-cadherin* protein in solid pseudopapillary tumors of the pancreas]. The activation of the *Wnt*/ $\beta$ -catenin pathway in SPT is highly correlated with *GLUL* expression, as would be expected of a *Wnt* target gene [118] and with high expression of proteins whose genes are located on chromosome 11q [119 – it shows that the accumulation of high expression of proteins whose genes are located on chromosome 11q is characteristic of SPT].

Recently, *Galectin-3* expression has been observed in both SPT and pancreatic normal ducts, suggesting the ductal origin of this tumor and representing a potentially useful marker to distinguish SPT from PET [120].

### Conclusion and Perspectives

The major clinical issues concerning pancreatic tumors are differential diagnosis and the related stratification of risk to guide management options. Although PDAC represents around 90% of all malignant tumors in the pancreas and has the worst prognosis, the correct recognition of other epithelial pancreatic tumors characterized by different behaviors is becoming increasingly relevant. The review of molecular studies highlights the correlation between the morphological classification of pancreatic neoplasms and distinct molecular profiles, suggesting the involvement of different molecular pathways that lead to the various subtypes of pancreatic neoplasms. IPMT and MCT share some molecular aspects with PDAC, representing probable precursor lesions, whereas SCT, PET, PACC, PBL and SPT are thoroughly distinct neoplasms with different pathogenetic and molecular mechanisms. Since different molecular pathways are associated with distinct patterns of molecular expression, future studies should also consider the correct classification of pancreatic epithelial tumors to identify novel diagnostic molecular tools and possible targets for innovative therapeutic strategies.

### Acknowledgements

This study was supported by a MolDiagPaca European Community FP6 grant (LSHB-CT-2006-018771) and Fondazione Cariverona, Italy.

## References

- 1 Klöppel G, Hruban RH, Longnecker DS, Adler G, Kern SE, Partanen TJ: Ductal adenocarcinoma of the pancreas; in Hamilton SR, Aaltonen LA (eds): Pathology and Genetics of Tumours of the Digestive System. WHO Classification of Tumours. Lyon, IARC Press, 2000.
- 2 Klöppel G et al: Classification of pancreatic neoplasms and their genetics; in Gress T, Neoptolemos J, Lemoine N, Real F (eds): Exocrine Pancreas Cancer. Ulm, The European Pancreatic Cancer-Research Cooperative, 2004.
- 3 Zamboni G, Klöppel G, Hruban RH, Longnecker DS, Adler G: Mucinous cystic neoplasms of the pancreas; in Hamilton SR, Aaltonen LA (eds): Pathology and Genetics of Tumours of the Digestive System. WHO Classification of Tumours. Lyon, IARC Press, 2000.
- 4 Terris B, Ponsot P, Paye F, Hammel P, Sauvanet A, Molas G, et al: Intraductal papillary mucinous tumors of the pancreas confined to secondary ducts show less aggressive pathologic features as compared with those involving the main pancreatic duct. *Am J Surg Pathol* 2000;24:1372–1377.
- 5 Sohn TA, Yeo CJ, Cameron JL, Hruban RH, Fukushima N, Campbell KA, et al: Intraductal papillary mucinous neoplasms of the pancreas: an updated experience. *Ann Surg* 2004;239:788–797; discussion 797–799.
- 6 Furukawa T, Klöppel G, Volkan Adsay N, Albores-Saavedra J, Fukushima N, Horii A, et al: Classification of types of intraductal papillary-mucinous neoplasm of the pancreas: a consensus study. *Virchows Arch* 2005;447:794–799.
- 7 Luttgies J, Zamboni G, Longnecker D, Klöppel G: The immunohistochemical mucin expression pattern distinguishes different types of intraductal papillary mucinous neoplasms of the pancreas and determines their relationship to mucinous noncystic carcinoma and ductal adenocarcinoma. *Am J Surg Pathol* 2001;25:942–948.
- 8 Adsay NV, Merati K, Basturk O, Iacobuzio-Donahue C, Levi E, Cheng JD, et al: Pathologically and biologically distinct types of epithelium in intraductal papillary mucinous neoplasms: delineation of an ‘intestinal’ pathway of carcinogenesis in the pancreas. *Am J Surg Pathol* 2004;28:839–848.
- 9 Miyasaka Y, Nagai E, Yamaguchi H, Fujii K, Inoue T, Ohuchida K, et al: The role of the DNA damage checkpoint pathway in intraductal papillary mucinous neoplasms of the pancreas. *Clin Cancer Res* 2007;13:4371–4377.
- 10 Furukawa T: Molecular genetics of intraductal papillary-mucinous neoplasms of the pancreas. *J Hepatobiliary Pancreat Surg* 2007;14:233–237.
- 11 Satoh K, Shimosegawa T, Moriizumi S, Koizumi M, Toyota T: K-ras mutation and p53 protein accumulation in intraductal mucin-hypersecreting neoplasms of the pancreas. *Pancreas* 1996;12:362–368.
- 12 Kitago M, Ueda M, Aiura K, Suzuki K, Hoshimoto S, Takahashi S, et al: Comparison of K-ras point mutation distributions in intraductal papillary-mucinous tumors and ductal adenocarcinoma of the pancreas. *Int J Cancer* 2004;110:177–182.
- 13 Sessa F, Solcia E, Capella C, Bonato M, Scarpa A, Zamboni G, et al: Intraductal papillary-mucinous tumours represent a distinct group of pancreatic neoplasms: an investigation of tumour cell differentiation and K-ras, p53 and c-erbB-2 abnormalities in 26 patients. *Virchows Arch* 1994;425:357–367.
- 14 Semba S, Moriya T, Kimura W, Yamakawa M: Phosphorylated Akt/PKB controls cell growth and apoptosis in intraductal papillary-mucinous tumor and invasive ductal adenocarcinoma of the pancreas. *Pancreas* 2003;26:250–257.
- 15 House MG, Guo M, Iacobuzio-Donahue C, Herman JG: Molecular progression of promoter methylation in intraductal papillary mucinous neoplasms (IPMN) of the pancreas. *Carcinogenesis* 2003;24:193–198.
- 16 Furukawa T, Fujisaki R, Yoshida Y, Kanai N, Sunamura M, Abe T, et al: Distinct progression pathways involving the dysfunction of DUSP6/MKP-3 in pancreatic intraepithelial neoplasia and intraductal papillary-mucinous neoplasms of the pancreas. *Mod Pathol* 2005;18:1034–1042.
- 17 Inoue H, Furukawa T, Sunamura M, Takeda K, Matsuno S, Horii A: Exclusion of SMAD4 mutation as an early genetic change in human pancreatic ductal tumorigenesis. *Genes Chromosomes Cancer* 2001;31:295–299.
- 18 Iacobuzio-Donahue CA, Klimstra DS, Adsay NV, Wilentz RE, Argani P, Sohn TA, et al: Dpc-4 protein is expressed in virtually all human intraductal papillary mucinous neoplasms of the pancreas: comparison with conventional ductal adenocarcinomas. *Am J Pathol* 2000;157:755–761.
- 19 Sato N, Rosty K, Jansen M, Fukushima N, Ueki T, Yeo CJ, et al: STK11/LKB1 Peutz-Jeghers gene inactivation in intraductal papillary-mucinous neoplasms of the pancreas. *Am J Pathol* 2001;159:2017–2022.
- 20 Xu S, Furukawa T, Kanai N, Sunamura M, Horii A: Abrogation of DUSP6 by hypermethylation in human pancreatic cancer. *J Hum Genet* 2005;50:159–167.
- 21 Sato N, Goggins M: Epigenetic alterations in intraductal papillary mucinous neoplasms of the pancreas. *J Hepatobiliary Pancreat Surg* 2006;13:280–285.
- 22 Terris B, Blaveri E, Crnogorac-Jurcovic T, Jones M, Missiaglia E, Ruzsniowski P, et al: Characterization of gene expression profiles in intraductal papillary-mucinous tumors of the pancreas. *Am J Pathol* 2002;160:1745–1754.
- 23 Sato N, Fukushima N, Maitra A, Iacobuzio-Donahue CA, van Heek NT, Cameron JL, et al: Gene expression profiling identifies genes associated with invasive intraductal papillary mucinous neoplasms of the pancreas. *Am J Pathol* 2004;164:903–914.
- 24 Sato N, Matsubayashi H, Abe T, Fukushima N, Goggins M: Epigenetic down-regulation of CDKN1C/p57KIP2 in pancreatic ductal neoplasms identified by gene expression profiling. *Clin Cancer Res* 2005;11:4681–4688.
- 25 Ohuchida K, Mizumoto K, Fujita H, Yamaguchi H, Konomi H, Nagai E, et al: Sonic hedgehog is an early developmental marker of intraductal papillary mucinous neoplasms: clinical implications of mRNA levels in pancreatic juice. *J Pathol* 2006;210:42–48.
- 26 Liu MS, Yang PY, Yeh TS: Sonic hedgehog signaling pathway in pancreatic cystic neoplasms and ductal adenocarcinoma. *Pancreas* 2007;34:340–346.
- 27 Jang KT, Lee KT, Lee JG, Choi SH, Heo JS, Choi DW, et al: Immunohistochemical expression of Sonic hedgehog in intraductal papillary mucinous tumor of the pancreas. *Appl Immunohistochem Mol Morphol* 2007;15:294–298.
- 28 Yamaguchi H, Inoue T, Eguchi T, Miyasaka Y, Ohuchida K, Mizumoto K, et al: Fascin overexpression in intraductal papillary mucinous neoplasms (adenomas, borderline neoplasms, and carcinomas) of the pancreas, correlated with increased histological grade. *Mod Pathol* 2007;20:552–561.
- 29 Schonleben F, Qiu W, Ciau NT, Ho DJ, Li X, Allendorf JD, et al: PIK3CA mutations in intraductal papillary mucinous neoplasm/carcinoma of the pancreas. *Clin Cancer Res* 2006;12:3851–3855.
- 30 Hashimoto Y, Murakami Y, Uemura K, Hayashidani Y, Sudo T, Ohge H, et al: Detection of human telomerase reverse transcriptase (hTERT) expression in tissue and pancreatic juice from pancreatic cancer. *Surgery* 2008;143:113–125.
- 31 Kanno A, Satoh K, Kimura K, Hirota M, Umino J, Masamune A, et al: The expression of MUC4 and MUC5AC is related to the biologic malignancy of intraductal papillary mucinous neoplasms of the pancreas. *Pancreas* 2006;33:391–396.
- 32 Zamboni G, Scarpa A, Bogina G, Iacono C, Bassi C, Talamini G, et al: Mucinous cystic tumors of the pancreas: clinicopathological features, prognosis, and relationship to other mucinous cystic tumors. *Am J Surg Pathol* 1999;23:410–422.

- 33 Wilentz RE, Albores-Saavedra J, Zahurak M, Talamini MA, Yeo CJ, Cameron JL, et al: Pathologic examination accurately predicts prognosis in mucinous cystic neoplasms of the pancreas. *Am J Surg Pathol* 1999;23:1320-1327.
- 34 Fukushima N, Fukayama M: Mucinous cystic neoplasms of the pancreas: pathology and molecular genetics. *J Hepatobiliary Pancreat Surg* 2007;14:238-242.
- 35 Luttgies J, Feyerabend B, Buchelt T, Pacena M, Kloppel G: The mucin profile of noninvasive and invasive mucinous cystic neoplasms of the pancreas. *Am J Surg Pathol* 2002;26:466-471.
- 36 Sawai H, Okada Y, Funahashi H, Matsuo Y, Tanaka M, Manabe T: Immunohistochemical analysis of molecular biological factors in intraductal papillary-mucinous tumors and mucinous cystic tumors of the pancreas. *Scand J Gastroenterol* 2004;39:1159-1165.
- 37 Jimenez RE, Warshaw AL, Z'Graggen K, Hartwig W, Taylor DZ, Compton CC, et al: Sequential accumulation of K-ras mutations and p53 overexpression in the progression of pancreatic mucinous cystic neoplasms to malignancy. *Ann Surg* 1999;230:501-509; discussion 509-511.
- 38 Kim SG, Wu TT, Lee JH, Yun YK, Issa JP, Hamilton SR, et al: Comparison of epigenetic and genetic alterations in mucinous cystic neoplasm and serous microcystic adenoma of pancreas. *Mod Pathol* 2003;16:1086-1094.
- 39 Gerdes B, Wild A, Wittenberg J, Barth P, Ramaswamy A, Kersting M, et al: Tumor-suppressing pathways in cystic pancreatic tumors. *Pancreas* 2003;26:42-48.
- 40 Izeradjene K, Combs C, Best M, Gopinathan A, Wagner A, Grady WM, et al: Kras(G12D) and Smad4/Dpc4 haploinsufficiency cooperate to induce mucinous cystic neoplasms and invasive adenocarcinoma of the pancreas. *Cancer Cell* 2007;11:229-243.
- 41 Fukushima N, Sato N, Yeo CJ, Cameron JL, et al: Characterization of gene expression in mucinous cystic neoplasms of the pancreas using oligonucleotide microarrays. *Oncogene* 2004;23:9042-9051.
- 42 Ishiguro H, Kato K, Kishimoto T, Nagai Y, Takahashi T, Sasano H, et al: Expression of steroidogenic enzymes by luteinizing cells in the ovarian-type stroma of a mucin-producing cystic tumour of the pancreas. *Histopathology* 2003;43:97-98.
- 43 Sorio C, Capelli P, Lissandrini D, Moore PS, Balzarini P, Falconi M, et al: Mucinous cystic carcinoma of the pancreas: a unique cell line and xenograft model of a preinvasive lesion. *Virchows Arch* 2005;446:239-245.
- 44 Capella C, Solcia E, et al: Serous cystic neoplasms of the pancreas; in Hamilton SR, Aaltonen LA (eds): *Pathology and Genetics of Tumours of the Digestive System*. WHO Classification of Tumours. Lyon, IARC Press, 2000.
- 45 Compton CC: Serous cystic tumors of the pancreas. *Semin Diagn Pathol* 2000;17:43-55.
- 46 George DH, Murphy F, Michalski R, Ulmer BG: Serous cystadenocarcinoma of the pancreas: a new entity? *Am J Surg Pathol* 1989;13:61-66.
- 47 Kosmahl M, Seada LS, Janig U, Harms D, Kloppel G: Solid-pseudopapillary tumor of the pancreas: its origin revisited. *Virchows Arch* 2000;436:473-480.
- 48 Kosmahl M, Wagner J, Peters K, Sipos B, Kloppel G: Serous cystic neoplasms of the pancreas: an immunohistochemical analysis revealing alpha-inhibin, neuron-specific enolase, and MUC6 as new markers. *Am J Surg Pathol* 2004;28:339-346.
- 49 Hoorens A, Prenzel K, Lemoine NR, Kloppel G: Undifferentiated carcinoma of the pancreas: analysis of intermediate filament profile and Ki-ras mutations provides evidence of a ductal origin. *J Pathol* 1998;185:53-60.
- 50 Ji Y, Wang XN, Lou WH, Sujie A, Tan YS, Jin DY: Serous cystic neoplasms of the pancreas: a clinicopathologic and immunohistochemical analysis. *Chin J Dig Dis* 2006;7:39-44.
- 51 Moore PS, Orlandini S, Zamboni G, Capelli P, Rigaud G, Falconi M, et al: Pancreatic tumours: molecular pathways implicated in ductal cancer are involved in ampullary but not in exocrine nonductal or endocrine tumorigenesis. *Br J Cancer* 2001;84:253-262.
- 52 Moore PS, Zamboni G, Brighenti A, Lissandrini D, Antonello D, Capelli P, et al: Molecular characterization of pancreatic serous microcystic adenomas: evidence for a tumor suppressor gene on chromosome 10q. *Am J Pathol* 2001;158:317-321.
- 53 Gumbs AA, Moore PS, Falconi M, Bassi C, Beghelli S, Modlin I, et al: Review of the clinical, histological, and molecular aspects of pancreatic endocrine neoplasms. *J Surg Oncol* 2002;81:45-53; discussion 54.
- 54 Klöppel G, Perren A, PU. H: The gastroenteropancreatic neuroendocrine cell system and its tumors. The WHO classification. *Ann N Y Acad Sci* 2004;1014:13-27.
- 55 Solcia E, Kloppel G, LH S: *Histological Typing of Endocrine Tumours*. New York, Springer, 2001.
- 56 Stott FJ, Bates S, James MC, McConnell BB, Starborg M, Brookes S, et al: The alternative product from the human CDKN2A locus, p14(ARF), participates in a regulatory feedback loop with p53 and MDM2. *EMBO J* 1998;17:5001-5014.
- 57 Muscarella P, Melvin WS, Fisher WE, Foor J, Ellison EC, Herman JG, et al: Genetic alterations in gastrinomas and nonfunctioning pancreatic neuroendocrine tumors: an analysis of p16/MTS1 tumor suppressor gene inactivation. *Cancer Res* 1998;58:237-240.
- 58 House MG, Schulick RD: Endocrine tumors of the pancreas. *Curr Opin Oncol* 2006;18:23-29.
- 59 Bartsch DK, Kersting M, Wild A, Ramaswamy A, Gerdes B, Schuermann M, et al: Low frequency of p16(INK4a) alterations in insulinomas. *Digestion* 2000;62:171-177.
- 60 Cupisti K, Hoppner W, Dotzenrath C, Simon D, Berndt I, Roher HD, et al: Lack of MEN1 gene mutations in 27 sporadic insulinomas. *Eur J Clin Invest* 2000;30:325-329.
- 61 Goebel SU, Heppner C, Burns AL, Marx SJ, Spiegel AM, Zhuang Z, et al: Genotype/phenotype correlation of multiple endocrine neoplasia type 1 gene mutations in sporadic gastrinomas. *J Clin Endocrinol Metab* 2000;85:116-123.
- 62 Moore PS, Missiaglia E, Antonello D, Zamo A, Zamboni G, Corleto V, et al: Role of disease-causing genes in sporadic pancreatic endocrine tumors: MEN1 and VHL. *Genes Chromosomes Cancer* 2001;32:177-181.
- 63 Wang EH, Ebrahimi SA, Wu AY, Kashefi C, Passaro E Jr, Sawicki MP: Mutation of the MEN1 gene in sporadic pancreatic endocrine tumors. *Cancer Res* 1998;58:4417-4420.
- 64 Zhuang Z, Vortmeyer AO, Pack S, Huang S, Pham TA, Wang C, et al: Somatic mutations of the MEN1 tumor suppressor gene in sporadic gastrinomas and insulinomas. *Cancer Res* 1997;57:4682-4686.
- 65 Rigaud G, Missiaglia E, Moore PS, Zamboni G, Falconi M, Talamini G, et al: High resolution allelotype of nonfunctional pancreatic endocrine tumors: identification of two molecular subgroups with clinical implications. *Cancer Res* 2001;61:285-292.
- 66 Perren A, Komminoth P, PU H: Molecular genetics of gastroenteropancreatic endocrine tumors. *Ann N Y Acad Sci* 2004;1014:199-208.
- 67 Nagano Y, Kim do H, Zhang L, White JA, Yao JC, Hamilton SR, et al: Allelic alterations in pancreatic endocrine tumors identified by genome-wide single nucleotide polymorphism analysis. *Endocr Relat Cancer* 2007;14:483-492.
- 68 Rigaud G, Moore PS, Zamboni G, Orlandini S, Taruscio D, Paradisi S, et al: Allelotype of pancreatic acinar cell carcinoma. *Int J Cancer* 2000;88:772-777.
- 69 Hahn SA, Seymour AB, Hoque AT, Schutte M, da Costa LT, Redston MS, et al: Allelotype of pancreatic adenocarcinoma using xenograft enrichment. *Cancer Res* 1995;55:4670-4675.
- 70 Chung DC, Brown SB, Graeme-Cook F, Tilbotson LG, Warshaw AL, Jensen RT, et al: Localization of putative tumor suppressor loci by genome-wide allelotyping in human pancreatic endocrine tumors. *Cancer Res* 1998;58:3706-3711.
- 71 Jonkers YM, Claessen SM, Perren A, Schmitt AM, Hofland LJ, de Herder W, et al: DNA copy number status is a powerful predictor of poor survival in endocrine pancreatic tumor patients. *Endocr Relat Cancer* 2007;14:769-779.

- 72 Capurso G, Lattimore S, Crnogorac-Jurcevic T, Panzuto F, Milione M, Bhakta V, et al: Gene expression profiles of progressive pancreatic endocrine tumours and their liver metastases reveal potential novel markers and therapeutic targets. *Endocr Relat Cancer* 2006;13:541-558.
- 73 Hansel DE, Rahman A, House M, Ashfaq R, Berg K, Yeo CJ, et al: Met proto-oncogene and insulin-like growth factor binding protein 3 overexpression correlates with metastatic ability in well-differentiated pancreatic endocrine neoplasms. *Clin Cancer Res* 2004;10:6152-6158.
- 74 Maitra A, Hansel DE, Argani P, Ashfaq R, Rahman A, Naji A, et al: Global expression analysis of well-differentiated pancreatic endocrine neoplasms using oligonucleotide microarrays. *Clin Cancer Res* 2003;9:5988-5995.
- 75 Couvelard A, Hu J, Steers G, O'Toole D, Sauvaget A, Belghiti J, et al: Identification of potential therapeutic targets by gene-expression profiling in pancreatic endocrine tumors. *Gastroenterology* 2006;131:1597-1610.
- 76 Roldo C, Missiaglia E, Hagan JP, Falconi M, Capelli P, Bersani S, et al: MicroRNA expression abnormalities in pancreatic endocrine and acinar tumors are associated with distinctive pathologic features and clinical behavior. *J Clin Oncol* 2006;24:4677-4684.
- 77 Di Florio A, Capurso G, Milione M, Panzuto F, Geremia R, Delle Fave G, et al: Src family kinase activity regulates adhesion, spreading and migration of pancreatic endocrine tumour cells. *Endocr Relat Cancer* 2007;14:111-124.
- 78 Hansel DE, Rahman A, Hermans J, de Krijger RR, Ashfaq R, Yeo CJ, et al: Liver metastases arising from well-differentiated pancreatic endocrine neoplasms demonstrate increased VEGF-C expression. *Mod Pathol* 2003;16:652-659.
- 79 Hansel DE, House MG, Ashfaq R, Rahman A, Yeo CJ, Maitra A: *MAGE1* is expressed by a subset of pancreatic endocrine neoplasms and associated lymph node and liver metastases. *Int J Gastrointest Cancer* 2003;33:141-147.
- 80 Karkkainen MJ, Petrova TV: Vascular endothelial growth factor receptors in the regulation of angiogenesis and lymphangiogenesis. *Oncogene* 2000;19:5598-5605.
- 81 Rahman A, Maitra A, Ashfaq R, Yeo CJ, Cameron JL, Hansel DE: Loss of p27 nuclear expression in a prognostically favorable subset of well-differentiated pancreatic endocrine neoplasms. *Am J Clin Pathol* 2003;120:685-690.
- 82 Lindberg D, Hessman O, Akerstrom G, Westin G: Cyclin-dependent kinase 4 (CDK4) expression in pancreatic endocrine tumors. *Neuroendocrinology* 2007;86:112-118.
- 83 Fjallskog ML, Hessman O, Eriksson B, Jansson ET: Upregulated expression of PDGF receptor beta in endocrine pancreatic tumors and metastases compared to normal endocrine pancreas. *Acta Oncol* 2007;46:741-746.
- 84 Borka K, Kaliszky P, Szabo E, Lotz G, Kupcsulik P, Schaff Z, et al: Claudin expression in pancreatic endocrine tumors as compared with ductal adenocarcinomas. *Virchows Arch* 2007;450:549-557.
- 85 Takahashi Y, Akishima-Fukasawa Y, Kobayashi N, Sano T, Kosuge T, Nimura Y, et al: Prognostic value of tumor architecture, tumor-associated vascular characteristics, and expression of angiogenic molecules in pancreatic endocrine tumors. *Clin Cancer Res* 2007;13:187-196.
- 86 Dalai I, Missiaglia E, Barbi S, Butturini G, Doglioni C, Falconi M, et al: Low expression of ARHI is associated with shorter progression-free survival in pancreatic endocrine tumors. *Neoplasia* 2007;9:181-183.
- 87 Missiaglia E, Moore PS, Williamson J, Lemoine NR, Falconi M, Zamboni G, et al: Sex chromosome anomalies in pancreatic endocrine tumors. *Int J Cancer* 2002;98:532-538.
- 88 Chang MC, Xiao S, Nose V: Clinicopathologic and immunohistochemical correlation in sporadic pancreatic endocrine tumors: possible roles of utrophin and cyclin D1 in malignant progression. *Hum Pathol* 2007;38:732-740.
- 89 Mourra N, Couvelard A, Tiret E, Olschwang S, Flejou JF: Clusterin is highly expressed in pancreatic endocrine tumours but not in solid pseudopapillary tumours. *Histopathology* 2007;50:331-337.
- 90 Ekeblad S, Lejonklou MH, Grimfjard P, Johansson T, Eriksson B, Grimelius L, et al: Co-expression of ghrelin and its receptor in pancreatic endocrine tumours. *Clin Endocrinol (Oxf)* 2007;66:115-122.
- 91 House MG, Herman JG, Guo MZ, Hooker CM, Schulick RD, Lillemoe KD, et al: Aberrant hypermethylation of tumor suppressor genes in pancreatic endocrine neoplasms. *Ann Surg* 2003;238:423-431; discussion 431-432.
- 92 Cecconi D, Donadelli M, Rinalducci S, Zolla L, Scupoli MT, Scarpa A, et al: Proteomic analysis of pancreatic endocrine tumor cell lines treated with the histone deacetylase inhibitor trichostatin A. *Proteomics* 2007;7:1644-1653.
- 93 Chen M, Rahman L, Voeller D, Kastanos E, Yang SX, Feigenbaum L, et al: Transgenic expression of human thymidylate synthase accelerates the development of hyperplasia and tumors in the endocrine pancreas. *Oncogene* 2007;26:4817-4824.
- 94 Klimstra DS, Heffess CS, Oertel JE, Rosai J: Acinar cell carcinoma of the pancreas. A clinicopathologic study of 28 cases. *Am J Surg Pathol* 1992;16:815-837.
- 95 Klimstra DS, Wenig BM, Adair CF, Heffess CS: Pancreatoblastoma. A clinicopathologic study and review of the literature. *Am J Surg Pathol* 1995;19:1371-1389.
- 96 Hoorens A, Gebhard F, Kraft K, Lemoine NR, Kloppel G: Pancreatoblastoma in an adult: its separation from acinar cell carcinoma. *Virchows Arch* 1994;424:485-490.
- 97 Pellegata NS, Sessa F, Renault B, Bonato M, Leone BE, Solcia E, et al: K-ras and p53 gene mutations in pancreatic cancer: ductal and nonductal tumors progress through different genetic lesions. *Cancer Res* 1994;54:1556-1560.
- 98 Cao D, Maitra A, Saavedra JA, Klimstra DS, Adsay NV, Hruban RH: Expression of novel markers of pancreatic ductal adenocarcinoma in pancreatic nonductal neoplasms: additional evidence of different genetic pathways. *Mod Pathol* 2005;18:752-761.
- 99 Lin YC, Lee PH, Yao YT, Hsiao JK, Sheu JC, Chen CH: Alpha-fetoprotein-producing pancreatic acinar cell carcinoma. *J Formos Med Assoc* 2007;106:669-672.
- 100 Kolb-van Harten P, Rosien U, Kloppel G, Layer P: Pancreatic acinar cell carcinoma with excessive alpha-fetoprotein expression. *Pancreatol* 2007;7:370-372.
- 101 Saif MW: Pancreatoblastoma. *JOP* 2007;8:55-63.
- 102 Abraham SC, Wu TT, Hruban RH, Lee JH, Yeo CJ, Conlon K, et al: Genetic and immunohistochemical analysis of pancreatic acinar cell carcinoma: frequent allelic loss on chromosome 11p and alterations in the APC/beta-catenin pathway. *Am J Pathol* 2002;160:953-962.
- 103 Abraham SC, Wu TT, Klimstra DS, Finn LS, Lee JH, Yeo CJ, et al: Distinctive molecular genetic alterations in sporadic and familial adenomatous polyposis-associated pancreatoblastomas: frequent alterations in the APC/beta-catenin pathway and chromosome 11p. *Am J Pathol* 2001;159:1619-1627.
- 104 Kerr NJ, Chun YH, Yun K, Heathcott RW, Reeve AE, Sullivan MJ: Pancreatoblastoma is associated with chromosome 11p loss of heterozygosity and IGF2 overexpression. *Med Pediatr Oncol* 2002;39:52-54.
- 105 Tanaka Y, Kato K, Notohara K, Nakatani Y, Miyake T, Ijiri R, et al: Significance of aberrant (cytoplasmic/nuclear) expression of beta-catenin in pancreatoblastoma. *J Pathol* 2003;199:185-190.
- 106 Nishimata S, Kato K, Tanaka M, Ijiri R, Toyoda Y, Kigasawa H, et al: Expression pattern of keratin subclasses in pancreatoblastoma with special emphasis on squamoid corpuscles. *Pathol Int* 2005;55:297-302.
- 107 Wong IH, Chan J, Wong J, Tam PK: Ubiquitous aberrant RASSF1A promoter methylation in childhood neoplasia. *Clin Cancer Res* 2004;10:994-1002.

- 108 Tang LH, Aydin H, Brennan MF, Klimstra DS: Clinically aggressive solid pseudopapillary tumors of the pancreas: a report of two cases with components of undifferentiated carcinoma and a comparative clinicopathologic analysis of 34 conventional cases. *Am J Surg Pathol* 2005;29:512–519.
- 109 Santini D, Poli F, Lega S: Solid-papillary tumors of the pancreas: histopathology. *JOP* 2006;7:131–136.
- 110 Papavramidis T, Papavramidis S: Solid pseudopapillary tumors of the pancreas: review of 718 patients reported in English literature. *J Am Coll Surg* 2005;200:965–972.
- 111 Abraham SC, Klimstra DS, Wilentz RE, Yeo CJ, Conlon K, Brennan M, et al: Solid-pseudopapillary tumors of the pancreas are genetically distinct from pancreatic ductal adenocarcinomas and almost always harbor beta-catenin mutations. *Am J Pathol* 2002;160:1361–1369.
- 112 Min Kim S, Sun CD, Park KC, Kim HG, Lee WJ, Choi SH: Accumulation of beta-catenin protein, mutations in exon-3 of the beta-catenin gene and a loss of heterozygosity of 5q22 in solid pseudopapillary tumor of the pancreas. *J Surg Oncol* 2006;94:418–425.
- 113 Tanaka Y, Kato K, Notohara K, Hojo H, Ijiri R, Miyake T, et al: Frequent beta-catenin mutation and cytoplasmic/nuclear accumulation in pancreatic solid-pseudopapillary neoplasm. *Cancer Res* 2001;61:8401–8404.
- 114 El-Bahrawy MA, Rowan A, Horncastle D, Tomlinson I, Theis BA, Russell RC, et al: E-cadherin/catenin complex status in solid pseudopapillary tumor of the pancreas. *Am J Surg Pathol* 2008;32:1–7.
- 115 Kim MJ, Jang SJ, Yu E: Loss of E-cadherin and cytoplasmic-nuclear expression of beta-catenin are the most useful immunoprofiles in the diagnosis of solid-pseudopapillary neoplasm of the pancreas. *Hum Pathol* 2008;39:251–258.
- 116 Tang WW, Stelter AA, French S, Shen S, Qiu S, Venegas R, et al: Loss of cell-adhesion molecule complexes in solid pseudopapillary tumor of pancreas. *Mod Pathol* 2007;20:509–513.
- 117 Serra S, Salahshor S, Fagih M, Niakosari F, Radhi JM, Chetty R: Nuclear expression of E-cadherin in solid pseudopapillary tumors of the pancreas. *JOP* 2007;8:296–303.
- 118 Audard V, Cavard C, Richa H, Infante M, Couvelard A, Sauvanet A, et al: Impaired E-cadherin expression and glutamine synthetase overexpression in solid pseudopapillary neoplasm of the pancreas. *Pancreas* 2008;36:80–83.
- 119 Tiemann K, Heitling U, Kosmahl M, Kloppe G: Solid pseudopapillary neoplasms of the pancreas show an interruption of the Wnt-signaling pathway and express gene products of 11q. *Mod Pathol* 2007;20:955–960.
- 120 Geers C, Moulin P, Gigot JF, Weynand B, Deprez P, Rahier J, et al: Solid and pseudopapillary tumor of the pancreas – review and new insights into pathogenesis. *Am J Surg Pathol* 2006;30:1243–1249.